

Insulin Resistance and Alzheimer's Disease: Molecular Links & Clinical Implications

Karen F. Neumann^{1,2}, Leonel Rojo^{1,3}, Leonardo P. Navarrete^{1,2}, Gonzalo Farías¹, Paula Reyes¹ and Ricardo B. Maccioni^{1,2,*}

¹Laboratory of Cellular and Molecular Neurosciences, University of Chile, ²International Center for Biomedicine (ICC), Santiago, Chile and ³Arturo Prat University, Iquique, Chile

Abstract: Hyperinsulinemia as well as type II diabetes mellitus are among the risk factors for Alzheimer's disease (AD). However, the molecular and cellular basis that link insulin resistance disorders and diabetes with AD are far from clear. Here, we discuss the potential molecular mechanisms that may explain the participation of these metabolic disorders in the pathogenesis of AD. The human brain uses glucose as a primary fuel; insulin secreted by the pancreas cross the blood-brain barrier (BBB), reaching neurons and glial cells, and exerts a region-specific effect on glucose metabolism. Glucose homeostasis is critical for energy generation, neuronal maintenance, neurogenesis, neurotransmitter regulation, cell survival and synaptic plasticity. It also plays a key role in cognitive function. In an insulin resistance condition, there is a reduced sensitivity to insulin resulting in hyperinsulinemia; this condition persists for several years before becoming full-blown diabetes. Toxic levels of insulin negatively influence neuronal function and survival, and elevation of peripheral insulin concentration acutely increases its cerebrospinal fluid (CSF) concentration. Peripheral hyperinsulinemia correlates with an abnormal removal of the amyloid beta peptide (A β) and an increase of tau hyperphosphorylation as a result of augmented cdk5 and GSK3 β activities. This leads to cellular cascades that trigger a neurodegenerative phenotype and decline in cognitive function. Chronic peripheral hyperinsulinemia results in a reduction of insulin transport across the BBB and a reduced insulin signaling in brain, altering all of insulin's actions, including its anti-apoptotic effect. However, the increase in brain insulin levels resulting from its peripheral administration at optimal doses has shown a cognition-enhancing effect in patient with AD. Some drugs utilized in type II diabetes mellitus reduce cognitive impairment associated with AD. The link between insulin resistance and neurodegeneration and AD, and the possible therapeutic targets in preventing the insulin-resistance disorders are analyzed.

Keywords: Insulin, Insulin Resistance, Hyperinsulinemia, Diabetes, Cognition, Neurodegeneration, Alzheimer's Disease.

A) GLUCOSE & INSULIN SIGNALING

1. Insulin and Insulin Receptor (IR)

Insulin is a small polypeptide, with a molecular weight of about 6,000 kDa, synthesized in significant quantities in β -cells of the pancreas. An increased level of glucose in blood stimulates β -cells in pancreas to secrete insulin by exocytosis, which diffuses into the islets capillary blood [1]. Insulin provides both short-term and long-term homeostatic signals, since it is secreted acutely in response to an increase in blood glucose and because plasma insulin levels are directly correlated with the degree of long term increase in body adiposity [2,3].

Insulin exerts its effect on glucose uptake in peripheral tissue by binding to the Insulin Receptor (IRs), a cell surface protein, which belongs to the family of tyrosine kinase receptors. Binding of insulin leads to a rapid autophosphorylation on several tyrosine residues, which provide docking sites for adaptor proteins, such as the insulin receptor substrate (IRS) proteins. Docking of adaptor proteins induces the activation of downstream pathways such as the lipid

kinase phosphatidylinositol 3-kinase (PI3K) and the mitogen activation protein kinase (MAPK) cascade [4,5]. PI3K is associated with almost all of the metabolic actions of insulin (refer to Fig. 1).

The effects of insulin on the central nervous system (CNS) are affected by its availability to this separate physiological compartment. The term BBB in its most restrictive sense refers to the vascular bed of the CNS, which is specially modified to prevent the unrestricted transfer of molecules between the blood and the extracellular fluid of the CNS [6]. The BBB plays a critical role in the transduction of signals between the CNS and peripheral tissues. It does so through several mechanisms, including the direct transport of peptides and regulatory proteins such as insulin [7].

There is solid evidence that insulin can cross the BBB by a saturable transport process mediated by the insulin receptor protein [6, 8–14]. This transporter is not uniformly distributed throughout the BBB [7]. There is also evidence of local insulin synthesis in brain [12, 14–16], although its function has not been elucidated. Therefore, the origin of brain insulin is a subject under investigation. IRs are present in the CNS, and were localized for the first time by ligand autoradiography and confirmed by immunohistochemistry and autoradiography [17–19]. CNS insulin receptors differ from their peripheral counterparts both in structure, function, and molecu-

*Address correspondence to this author at the International Center for Biomedicine, Providencia 455, Dept. 303, Providencia, Santiago, Chile; E-mail: rmaccion@manquehue.net

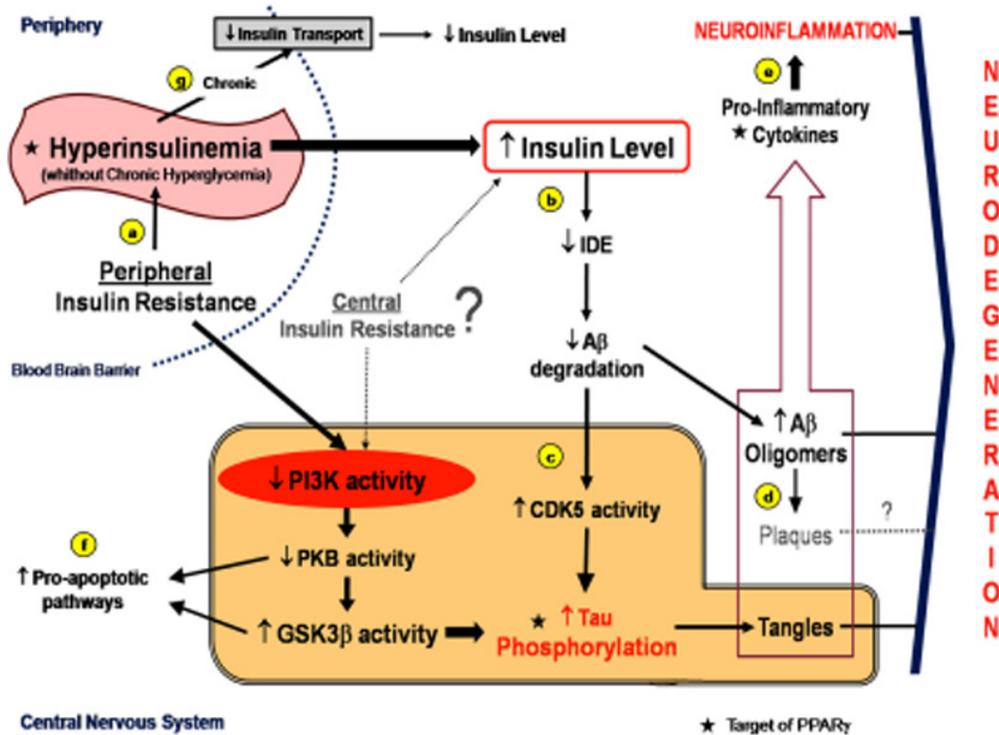


Fig. (1). Schematic representation of molecular pathways linking insulin resistance and neurodegeneration, and therapeutic target of PPAR γ ligands.

(a) Peripheral insulin resistance results in periods of hyperinsulinemia to overcome the hyperglycemia. (b) Hyperinsulinemia, with the possible contribution of central insulin resistance, leads to increased insulin levels in the CNS, that kidnaps IDE resulting in a decrease of A β degradation, which upregulates cdk5 with consequent tau hyperphosphorylation, in the meantime that increases A β oligomers. (c) Peripheral insulin resistance is associated with decreased insulin signaling in neuron, that also can be a result of central insulin resistance, with decreased PI3K and PKB activity, leading to an increased GSK3 β activity. Cdk5 and GSK3 β activities increase tau phosphorylation and tangles formation. (d) A β oligomers are more toxic than Senile Plaques, they are one of the responsible elements for activation of the innate immune response in the glia. (e) These oligomers are among factors that lead to Neuroinflammation. (f) Downregulation of PKB activity and a high GSK3 β activity are also related with pro-apoptotic pathways that result in neurodegeneration. (g) Thus, chronic hyperinsulinemia (without hyperglycemia) results in down-regulation of insulin transporters in the blood brain barrier and decreases the levels of insulin in the CNS.

lar weight [20]. IRs are widely distributed in the brain with the highest concentration in the olfactory bulb, hypothalamus, cerebral cortex, cerebellum and hippocampus. They are expressed in all regions of brain in both neurons and glial cells. IRs are distributed throughout the neuronal processes and the cell body, but are concentrated in the synaptic endings [4, 21–24].

The signaling and biological effects of the insulin have been widely studied mainly in the “classical” insulin target tissues, e.g. liver, fat and skeletal muscle, with respect to glucose uptake, regulation of cell proliferation, gene expression and the suppression of hepatic glucose production [4]. However, insulin plays many roles within the CNS. It has been shown that some of the CNS effects of insulin are the opposite to those exerted in peripheral tissues. In particular, CNS insulin increases glucose and inhibits feeding, whereas serum insulin decreases glucose and increases feeding [14,25]. Thus, to some extent, insulin acts as its own counterregulatory hormone, with CNS insulin producing features of insulin resistance [6]. Thus, in the CNS, insulin participates in the regulation of feeding behavior and energy homeostasis, neuronal maintenance, neurogenesis, and neurotransmitter regulation. In addition, it has a role on cognitive

functions as supported by neuronal activity, and in the control of aging-related processes [26].

2. Glucose, Insulin and Cognition

The brain consumes metabolic energy disproportionate to its size. It uses glucose (primary fuel), largely through oxidative metabolism. Glucose deprivation leads to coma, seizures, and produces potentially permanent brain damage. Glucose is continually supplied from cerebral blood flow and must be transported into the brain through the endothelial cells that form the BBB to reach neurons and glial cells by facilitated diffusion. Carriers of glucose are the glucose transport proteins (GLUT), which allow glucose entry into individual cells (neurons and glial cells) [27]. Animal models show that the cerebral blood flux depends on glucose levels in the blood stream; there is a compensatory mechanism to maintain adequate delivery of glucose fuel to the brain [28,29].

Evidence suggests regionally specific effects of insulin on brain glucose metabolism. Insulin does not seem to influence basal cerebral glucose metabolism or transport of glucose into the brain [30–35]. Insulin affects the use of glucose

in specific regions of the brain, most likely by selective distribution of insulin-sensitive GLUT isoforms, which overlap with the distribution of insulin and IRs in the brain [36–39]. Insulin, through the IRs localized in the hypothalamus, contributes to the regulation of food intake and energy homeostasis and leads to changes in body weight by anorexigenic or orexigenic effects produced by increasing or decreasing levels of insulin in brain respectively. Moreover, insulin in the CNS modifies peripheral glucose metabolism by increasing insulin sensitivity in peripheral tissue [10,40,41].

In brain, insulin is not a major regulator of glucose metabolism. *In vitro* studies showed that insulin regulates glucose uptake of glial cells, but did not influence neuronal glucose uptake [42,43]. Insulin can influence neurons directly by mechanisms unrelated to modulation of glucose uptake. Neurotransmitter release, neuronal-outgrowth, tubulin activity, neuronal survival and synaptic plasticity are all directly modulated by insulin [5, 44–46]

Studies in human and animal models have shown that an increase in brain insulin has a cognition-enhancing effect, independently of changes in peripheral glucose [47–51]. Moreover, there is cumulative evidence to support the effects of insulin and IRs on cognition, mediated by a modulatory role in learning and memory processes [1,4,5,26,52]. Insulin also modulates CNS concentration of neurotransmitters associated with important roles in cognition such as acetylcholine, norepinephrine and dopamine [53,54]. It has been shown that, in an early stage of memory formation, an alteration of gene expression of IRs in the rat hippocampus in response to learning experiences occurs [55]. Taken together, all these findings suggest that insulin may influence normal memory function (Figs. 1 and 2).

The biological basis of learning and memory processes resides in synaptic strength, where insulin signaling plays a modulator role on synaptic long-term potentiation (LTP) and long-term depression (LTD), two opposite forms of activity-dependent synaptic modifications. Insulin signaling modulates synaptic plasticity by: 1) Promoting the recruitment of GABA receptors on post-synaptic membranes; 2) Influencing NMDA receptor conductance (neuronal Ca^{2+} influx); and 3) Regulating AMPA receptor cycling [56–62].

B) INSULIN RESISTANCE

1. Insulin Resistance

Reduced sensitivity to insulin by the main target organs (liver, fat, muscle) is known as Insulin Resistance, where there is an elevated level of insulin in the bloodstream (hyperinsulinemia). This reduced sensitivity also results in impaired response to oral glucose (referred to as Impaired Glucose Tolerance), where it takes longer to restore normal glucose levels after eating. Peripheral Insulin Resistance is known to be the major contributor to the progression to hyperglycemia and Type II Diabetes Mellitus, since the pancreas can not secrete enough insulin to overcome the insulin resistance and prevent these events. Hyperinsulinemia without hyperglycemia is an indication of Insulin Resistance, a pre-diabetic condition. There is evidence of an increased risk of cognitive decline and neurodegeneration in populations with peripheral insulin resistance (hyperinsulinemia without hyperglycemia) [63–66].

2. Hyperinsulinemia/Hyperglycemia in the Brain

Alterations in circulating glucose levels can negatively affect the CNS because neurons have a consistently high glucose demand. Neuronal glucose uptake depends on extracellular glucose concentration, but chronic hyperglycemia results in cellular damage (glucose neurotoxicity). Therefore, a tight metabolic control is very important [67–69]. Studies on hyperglycemic rodents have shown cognitive impairment in addition to functional and structural alterations in the brain [68].

A growing body of evidence suggests that peripheral insulin abnormalities increase the risk for memory loss and neurodegenerative disorders such as AD, but acute and chronic hyperinsulinemia have opposing effects on memory performance (Fig. 2). A possible explanation for that is discussed below. Chronic hyperinsulinemia has a negative influence on memory, since type II Diabetes Mellitus has been associated with long-term impairment in cognitive function in humans and animal model studies. On the other hand, acute increases of peripheral or brain insulin have an enhanced memory performance effect [47,49, 63, 70–73]. Pre-diabetic conditions with hyperinsulinemia but not chronic hyperglycemia may persist for many years before progression to type II Diabetes Mellitus. Hyperinsulinemia exposes the cells (including neurons) to unphysiologically high levels of insulin for a long period of time. Studies have shown that high concentrations of insulin affect the function and survival of neurons in culture by sensitizing them to toxin and stress-induced insults [74].

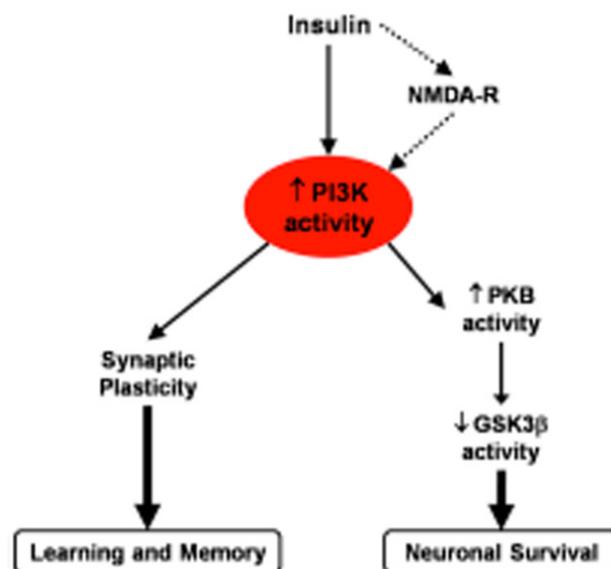


Fig. (2). Schematic representation of the relationships between insulin signaling, learning, memory and neuronal survival.

Acute high levels of insulin in the CNS have been related with the improvement of cognitive function and Neuronal Survival by a pathway that it involves the NMDA-R, PI3K and PKB activity and inactivation of GSK3 β .

Moreover, a relationship between glycosylation of tau proteins, its aggregation ability and the neurodegenerative phenotype has been revealed [75]. Peripheral injection of

high doses of insulin in mice caused a rapid and dose-dependent increase in tau phosphorylation in the CNS [76]. *In vitro* studies indicate that insulin modulates the levels of A β peptide by promoting the release of intracellular A β . Physiological insulin levels promote A β clearance by peptide degradation, a mechanism that involves insulin degrading enzyme (IDE) activity (detailed below in section III.1) [77–79]. A low level of insulin in brain reduces A β release from intracellular to extracellular compartments and high levels reduce A β degradation in the extracellular compartment [80–84].

Impaired verbal memory has been reported in individuals with hyperinsulinemia and not chronic hyperglycemia [85]. Epidemiological studies have indicated that hyperinsulinemia, independent of glucose levels, constitutes one of the risk factors for dementia in the type II diabetes mellitus population as well as in the non-diabetic population, since the raising of peripheral insulin concentration acutely increases its concentration in the CSF and brain [63,65,70,86]. Prolonged peripheral hyperinsulinemia downregulates BBB functions and the IR activity and reduces insulin transport into the brain [86, 87]. Thus, hyperinsulinemia during the development of type II diabetes mellitus is neurotoxic. Development of these complications depend on the duration of a diabetic condition, upregulation of circulating glucose, glycosylated proteins, etc., and the quality of metabolic control [67].

Insulin resistance, hyperinsulinemia and type II Diabetes Mellitus are associated with elevated inflammatory markers and increased risk for AD [63,65,88–90] (Fig. 1). In adults with type II Diabetes Mellitus and impaired glucose tolerance, an abnormal level of soluble TNF-R1 (death receptor domain) and TNF-R2 (cell survival) has been reported. Increased levels of TNF-R1 and decreased levels of TNF-R2 have also been observed in AD brain [91–93].

3. Impaired Glucose Uptake and Metabolism in the AD Brain

Studies using positron emission tomography (PET) have demonstrated that glucose metabolism is reduced markedly in the cerebral cortex in early stage AD and in the mild cognitive impairment (MCI) that is believed to be a precursor of AD, and that the reduction parallels the worsening of dementia symptoms, establishing that glucose uptake and metabolism are impaired in AD brain [94]. However, the causes of the impairment of glucose uptake/metabolism in AD brain are not well understood. Several findings suggest that this impairment is a cause, rather than a consequence, of neurodegeneration. The impaired cerebral glucose consumption is more severe than the impaired oxygen consumption in AD, suggesting that the former is not the result of the latter.

The brain is highly dependent upon glucose as a source of energy. The impaired glucose uptake/metabolism might lead to deficient synaptic activity and cellular homeostasis, as these are very sensitive to energy deficiency. Impaired glucose uptake/metabolism also causes reduced formation of acetyl-CoA and, consequently, affects the synthesis of acetylcholine [95]. Deficient activity of the cholinergic system is one of the most significant biochemical deficiencies in AD pathology, discovered decades ago [96], and is the basis of

the first generation of AD drugs, the cholinesterase inhibitors donepezil, galantamine and rivastigmine. The reduced cellular availability of acetyl-CoA may also cause decreased formation of both intracellular cholesterol and neurosteroids, which are the main lipid components of cell membranes. Membrane abnormalities have been observed in AD brains.

The A β peptide (or A β oligomers) has been shown to cause decreased glucose uptake/metabolism [97]. Furthermore, apolipoprotein E4 (apoE4) and diabetes mellitus are known to be major risk factors for AD [98]. Individuals with apoE4 alleles have lower brain glucose metabolism than those carrying apoE2 or apoE3 alleles. Diabetes patients are characterized by deficient glucose uptake/metabolism. Hence, it can be speculated that apoE4 genotype together with diabetes may increase the risk for AD by impairment in brain glucose uptake/metabolism.

C) LINK BETWEEN INSULIN RESISTANCE AND ALZHEIMER DISEASE

1. Relationships between Insulin Resistance and AD Pathogenesis

Abnormalities in insulin metabolism, characteristic of type II diabetes, are among the major factors thought to mechanistically influence the onset of AD. These abnormalities are thought to play a role in AD via their influence on the synthesis and degradation of A β and as a consequence of the cascade of neuronal alterations resulting from the effects of danger/alarm signals from oligomeric amyloid species (Fig. 1) [99]. Additionally, recent studies have indicated that certain signal transduction pathways downstream of the insulin receptor may also promote the generation of A β peptides by modulating the cleavage of the parent A β precursor protein (A β PP) at the γ -secretase site, a cleavage site necessary for A β amyloidogenicity [100].

Although this evidence tentatively suggests that type II Diabetes Mellitus might play an important role in AD through mechanisms that involve A β peptide generation, alternative studies suggest that insulin may also provoke amyloid accumulation by limiting A β degradation via direct competition for the IDE. Another major substrate of IDE is the A β peptide. The IDE degrades monomeric but not the oligomeric A β peptide [77,78]. IDE is a zinc-metallo-peptidase that preferentially cleaves proteins with a propensity to form β -pleated sheet-rich amyloid fibrils, such as A β peptides. This relationship of IDE with A β is supported by recent evidence indicating that IDE activity in the brain is negatively correlated with A β content, and that IDE expression is decreased in the AD brain [83]. Insulin regulates the levels of IDE. Hyperinsulinemia reduces A β degradation by reduction of IDE levels and binding competition (Fig. 1) [80–84].

It has been reported that A β 40 and A β 42 reduce insulin binding and insulin receptor autophosphorylation. The reduction in this binding seems to be caused by a decrease in the affinity of insulin to the insulin receptor. This suggests that A β is a direct competitive inhibitor of insulin binding and action [101], an aspect that demands further investigation.

The strikingly reduced CNS expression of genes encoding insulin, IGF-I, and IGF-II, as well as the insulin and IGF-I receptors in AD led to some authors to suggest that AD may represent a neuro-endocrine disorder that resembles diabetes mellitus [102]. In addition, the same researchers demonstrated that alterations insulin levels and insulin-like growth factor expression and deterioration of insulin function with the course of AD progression, were linked with an acetylcholine decrease in the brain [103]. There is evidence supporting the notion that high plasma insulin levels and peripheral insulin resistance affect A β 42 levels, inflammation in the CNS and cognitive performance of individuals [104]. From such evidence, a model can be constructed describing how this metabolic profile contributes to the pathogenesis of AD. There are several etiological factors leading to the final common expression in the AD pathology [103].

Insulin plays an important role in memory and brain function in general. Peripheral hyperinsulinemia and insulin resistance induce a number of deleterious effects in the CNS that interfere with these functions, in a manner that is exacerbated by obesity and aging. In particular, effects on A β regulation and neuroinflammation are potential culprits in promoting aging-related memory impairment in some cases of AD (see Fig. 1). This possibility has obvious relevance for adults with type II Diabetes Mellitus. However, it is worth noting that hyperinsulinemia and insulin resistance afflict many non-diabetic adults with conditions such as obesity, impaired glucose tolerance, cardiovascular disease, and hypertension [105–107].

Indeed, in recent years, cumulative evidence has been gained on the involvement of alteration in neuronal lipoproteins activity, as well as a role of cholesterol and other lipids in the pathogenesis of this neurodegenerative disorder. In relation to hypercholesterolemia, several reports have shown that elevated serum cholesterol levels and elevated levels of A β are linked with AD risk. Cholesterol influences the activity of the enzymes involved in the metabolism of the amyloid precursor protein and in the production of A β , but the mechanism by which cholesterol affects A β production and metabolism is not fully understood [108].

2. Metabolomics of Insulin in the Context of Neuronal Survival and AD Onset

To understand the link between insulin, insulin resistance, neuronal survival and AD onset, it is important identify the link between the key molecules involved in the intracellular pathways utilized by insulin to exert its effect. Insulin resistance in the periphery produces acute episodes of hyperinsulinemia without chronic hyperglycemia. High levels of insulin in plasma are correlated with high levels of insulin in brain (Fig. 1) [86], leading to neurotoxic effects [74].

Studies in transgenic mouse models of AD have shown that diet-induced peripheral insulin resistance promotes amyloidosis suggesting that peripheral insulin resistance can influence A β production in the brain [84,109]. These findings in association of a reduced basal signaling of insulin in cortex with an increased degree of AD neuropathology argue that peripheral insulin resistance promotes neuronal insulin

resistance when genetic background predisposes to AD. However, further studies are necessary to clarify this.

Insulin binding to the IRs leads to autophosphorylation of the IRs which initiates several signaling cascades. One of these is the lipid kinase phosphatidylinositol 3-kinase (PI3K) cascade, which is associated with almost all of the metabolic action of insulin. PI3K activation leads to the activation of Protein Kinase B (PKB or Akt) and Glycogen Synthase Kinase 3 β (GSK3 β). Activation of PKB inactivates GSK3 β by phosphorylation [110,111]. The neuroprotective effect of IGF-1 results from activation of PKB [112]. Expression of PKB protects neurons against toxin-induced death and protects PC12 cells against A β peptide-induced death [113–115].

Cole and coworkers have reviewed the molecular link between insulin action, diabetes and AD [5]. Insulin resistance in the periphery produces hyperinsulinemia, while in the brain it decreases IDE activity. The effects of IDE include abnormal A β removal and plaque formation, increased cdk5 activity, and increased activity of GSK3 β (Fig. 1). Dysregulation of cdk5 is a major molecular event in the pathway to neurodegeneration [116–118]. GSK3 β activity has been implicated in the pathology of AD in different ways. AD brain exhibits a dysregulated expression of this kinase as well as changes in its activity [119,120], leading to hyperphosphorylation of tau and tangle formation. Several studies have shown that GSK3 β activity is required for induction of neuronal apoptosis (refer to Fig. 1), while inhibition of GSK3 β promotes neuronal survival (illustrated schematically in Fig. 2) (reviewed by Cole *et al.* 2007 [5]). Insulin signaling induces the phosphorylation and inhibition of GSK3 β [110].

All of the negative effects of hyperinsulinemia in the CNS and their associated functions could be exacerbated or similarly produced by a central insulin resistance in conjunction with the peripheral insulin resistance or secondary to it. This is supported by the evidence of central molecular insulin resistance in a mouse model of AD with induced peripheral insulin resistance, where a reduced basal signaling (reduced phosphorylation of IR, PKB and GSK3, as well as decreased PI3K activity) was shown in the cerebral cortex (Fig. 1) [84,109].

Studies of insulin action in brain are focused on the basic effects of insulin signaling. Insulin acts like a neurotrophic factor, since it promotes neuronal survival [46,121]. Studies *in vitro* of intracellular pathways utilized by insulin for synaptic plasticity have identified a link to neuronal protection against cell death [46,121]. Intracellular pathways utilized by insulin to influence synaptic plasticity and neuronal survival converge on the PI3K pathway (Fig. 2) (reviewed by van der Heide *et al.* 2006 [1]). The increased catalytic activity of PI3K results in the phosphorylation and activation of anti-apoptotic substrates [1]. PKB is of major importance in mediating the effects of PI3K in neuronal survival, and *in vitro* and *in vivo* studies have shown that activated PKB protects against apoptosis. Dominant negative PKB does not [122–124]. Studies on insulin-facilitated LTP and LTD induction show that this process is completely NMDA receptor-dependent [62]. PI3K activity is required for LTP maintenance in the hippocampal CA1 region [125]. An increased

gene expression of IR in the rat hippocampus in response to learning experiences has been shown [55]. All together, the effect of insulin on neuronal survival and its effects on NMDA-dependent synaptic plasticity, require similar intracellular signaling routes (schematically illustrated in Fig. 2).

Clinical Studies on Insulin Resistance and AD

In the last five years a growing body of evidence has been developed suggesting that glucose metabolism is associated with the pathogenesis of AD, age-related cognitive decline and neuroinflammation [90,126–128]. Intravenous insulin infusion in healthy, euglycemic older subjects (a procedure known as hyperinsulinemic-euglycemic clamp) induces a facilitation of memory while it increases cerebrospinal fluid levels of $A\beta_{1-42}$ [126]. Insulin mediated memory facilitation is less marked at supraoptimal levels of insulin and in subjects with AD the hyperinsulinemic facilitation of memory is obtained at higher concentrations of insulin when compared with normal subjects. This phenomenon is more clear in patients who are not carriers of the APOE- $\epsilon 4$ gene [51].

The clinical effects of insulin and/or glucose administration on memory in patients with AD have been extensively reviewed [128,129]. As a result, it has been suggested that moderate administration of both glucose and insulin could improve memory in patients. These observations appear to be in disagreement with Gispén and Biessels [68] who described that both chronic hyperglycemia and chronic hyperinsulinemia are associated with accelerated cognitive decline in the elderly. This apparent discrepancy may be explained in part by the mechanisms underlying spontaneous chronic hyperglycemia and chronic hyperinsulinemia during insulin resistance. Hoyer [130] has comprehensively addressed this issue in his recent review. He classified AD as an “insulin resistant brain state”. This interesting hypothesis is supported by a recent clinical trial performed by Reger *et al.* [73] where the administration of intranasal insulin improved cognitive performance in elderly patients. In this pilot study, the authors administered 20 IU BID intranasal insulin to 13 patients and compared them with 12 controls receiving placebo. They found no changes in plasma levels of glucose and insulin with the treatment. However the insulin-treated group retained more verbal information after a delay and showed improved attention and functional status. Insulin treatment also increased the $A\beta_{1-40}/A\beta_{1-42}$ ratio in plasma. Previous studies by the same author [131] demonstrated that this effect was restricted to APOE- $\epsilon 4$ negative subjects.

Modulation of the response to insulin by the APOE genotype has led to speculation regarding whether APOE- $\epsilon 4$ is an independent risk factor for AD, so that people carrying this allele do not have a greater risk of being insulin resistant than the general population. In the other hand, people that are insulin resistant are at risk of developing AD regardless of their APOE- $\epsilon 4$ status [128]. The evidence that favors the role of insulin resistance in AD has opened new perspectives for treatment options. The family of antidiabetic Thiazolidinedione drugs function as agonists of the nuclear receptor peroxisome proliferator-activated receptor (PPAR)- γ and have been shown to improve sensitivity to insulin by decreasing circulating insulin and increasing insulin medi-

ated glucose uptake. In addition to these functions, they also have been demonstrated to diminish the levels of multiple inflammatory mediators (Fig. 1) (reviewed by Jiang, 2008, Rojo *et al.*, 2008 [90,133]). Studies in cellular models have shown that PPAR γ agonists down-regulate A β peptide deposition that occurs in AD, although the mechanisms of this phenomenon demand further investigation [133–135]. Troglitazone, a thiazolidinedione, significantly reduced phosphorylation of tau protein at Ser202 and Ser396/404, residues phosphorylated in early and later stages of neurofibrillary tangle accumulation in AD and other neurodegenerative disorders (Fig. 1, PPAR γ ligands) [136].

Interestingly, the PPAR γ Pro12Ala polymorphism, which is associated with an increased risk of type II diabetes [137], has been found to influence plasma 24S-hydroxycholesterol/cholesterol ratio in AD patients. This fact is important considering that elevated cholesterol in blood is a risk factor for AD and 24S-hydroxycholesterol is the major product of brain cholesterol metabolism and is released into the blood stream [108]. At present pioglitazone and rosiglitazone are the two thiazolidinediones that are available for clinical use. Both share the adverse side-effect of generating edema and potentially heart failure [138]. A recent meta-analysis [139] revealed a possible increased risk of myocardial infarction and a borderline increase in the risk of death from cardiovascular causes in patients receiving rosiglitazone. However, the significance of the data has been debated [140]. Thus, rosiglitazone as a possible therapy for AD is an active field of clinical investigation.

Rosiglitazone has been shown to preserve memory function in AD patients when compared with a placebo-assigned group. In a double blind trial, 20 mild AD or MCI patients who received 4 mg of rosiglitazone for 6 months showed improvements in memory and selective attention, associated with less reduction in plasma A β -40 and A β -42 when compared to 10 control patients [129]. This stabilization of plasma A β could be of importance, considering that plasma A β_{1-42} decreases with AD progression, as previously described [82]. Another study on 511 AD patients treated with 2 mg, 4 mg or 8 mg of rosiglitazone or placebo found that there is an improvement in cognitive function when evaluated by the Alzheimer’s Disease Assessment Scale-cognitive (ADAS-cog) for patients treated with 8 mg of rosiglitazone, and that this result is restricted to those patients who are not carriers of APOE- $\epsilon 4$ [141]. This modulation of the effect of rosiglitazone by the APOE genotype strongly resembles what has been observed for insulin and support the recommendation for APOE genotyping of patients at risk of cognitive decline [142]. Even though this evidence is still preliminary, it supports the role of PPAR γ ligands with their dual action over insulin resistance and on neuroinflammation as a novel strategy for the treatment of cognitive decline associated with AD and stress the importance of further studies in this perspective.

ACKNOWLEDGMENTS

Research has been supported by a grant from Fondecyt 1080254 and a grant from the Alzheimer’s Association, U.S.A. Authors are grateful to Sarah Murray MD. for carefully revising the text of this paper.

REFERENCES

- [1] Van der Heide LP, Ramakers GM and Smidt MP. Insulin signaling in the central nervous system: Learning to survive. *Prog Neurobiol* 79: 205–221 (2006).
- [2] Bagdade JD, Bierman EL and Porte Jr D. The Significance of Basal Insulin Levels in the Evaluation of the Insulin Response to Glucose in Diabetic and Nondiabetic Subjects. *J Clin Invest* 46: 1549–1557 (1967).
- [3] Polonsky KS, Given BD, Hirsch L, Shapiro ET, Tillil H, Beebe C, *et al.* Quantitative Study of Insulin Secretion and Clearance in Normal and Obese Subjects. *J Clin Invest* 81:435–441 (1987).
- [4] Plum L, Schubert M and Brüning JC. The role of insulin receptor signaling in the brain. *Trends Endocrinol Metab* 16: 59–65 (2005).
- [5] Cole AR, Astell A, Green C and Sutherland C. Molecular connexions between dementia and diabetes. *Neurosci Biobehav Rev* 31(7): 1046–1063 (2007).
- [6] Banks WA. The source of cerebral insulin. *Eur J Pharmacol* 490: 5–12 (2004).
- [7] Banks WA. Blood-brain barrier and energy balance. *Obesity (Silver Spring)* 14 Suppl 5: 234S–237S (2006).
- [8] Margolis RU and Altszuler N. Insulin in the cerebrospinal fluid. *Nature* 215: 1375–1376 (1967).
- [9] Steffens AB, Scheurink AJ, Porte D Jr and Woods SC. Penetration of peripheral glucose and insulin into cerebrospinal fluid in rats. *Am J Physiol* 255: R200–R204 (1988).
- [10] Schwartz MW, Figlewicz DP, Baskin DG, Woods SC and Porte D Jr. Insulin in the brain: a hormonal regulator of energy balance. *Endocr Rev* 13: 387–414 (1992).
- [11] Baura GD, Foster DM, Porte D Jr, Kahn SE, Bergman RN, Cobelli C, *et al.* Saturable transport of insulin from plasma into the central nervous system of dogs *in vivo*. A mechanism for regulated insulin delivery to the brain. *J Clin Invest* 92: 1824–1830 (1993).
- [12] Banks WA, Jaspan JB and Kastin AJ. Selective, physiological transport of insulin across the blood-brain barrier: novel demonstration by species-specific radioimmunoassay. *Peptides* 18:1257–1262 (1997).
- [13] Banks WA, Jaspan JB, Huang W and Kastin AJ. Transport of insulin across the blood-brain barrier: saturability at euglycemic doses of insulin. *Peptides* 18 (9):1423–1429 (1997).
- [14] Woods SC, Seeley RJ, Baskin DG and Schwartz MW. Insulin and blood-brain barrier. *Curr Pharm Des* 9(10): 795–800 (2003).
- [15] Devaskar SU, Giddings SJ, Rajakumar PA, Carnaghi LR, Menon RK and Zahm DS. Insulin gene expression and insulin synthesis in mammalian neuronal cells. *J Biol Chem* 269: 8445–8454 (1994).
- [16] Rulifson EJ, Kim SK and Nusse R. Ablation of insulin-producing neurons in flies: growth and diabetic phenotypes. *Science* 296: 1118–1120 (2002).
- [17] Havrankova J and Roth J. Insulin receptors are widely distributed in central nervous-system of rat. *Nature* 212: 827–829 (1978).
- [18] van Houten M, Posner BI, Kopriwa BM and Brawer JR. Insulin-binding sites in the rat brain: *in vivo* localization to the circumventricular organs by quantitative radioautography. *Endocrinology* 105(3): 666–673 (1979).
- [19] Baskin DG, Woods SC, West DB, van Houten M, Posner BI, Dorsa DM, *et al.* Immunocytochemical detection of insulin in rat hypothalamus and its possible uptake from cerebrospinal fluid. *Endocrinology* 113: 1818–1825 (1983).
- [20] Heidenreich KA, Zahniser NR, Berhanu P, Brandenburg D and Olefsky JM. Structural differences between insulin receptors in the brain and peripheral target tissues. *J Biol Chem* 258: 8527–8530 (1983).
- [21] Havrankova J, Brownstein M and Roth J. Insulin and insulin-receptors in rodent brain. *Diabetologia* 20: 26873 (1981).
- [22] Wozniak M, Rydzewski B, Baker SP and Raizada M. The cellular and physiological actions of insulin in the central nervous system. *Neurochem Int* 22: 1–10 (1993).
- [23] Unger JW and Betz M. Insulin receptors and signal transduction proteins in the hypothalamo-hypophyseal system: a review on morphological findings and functional implications. *Histol Histopathol* 13: 1215–1224 (1998).
- [24] Abbott MA, Wells DG and Fallon JR. The insulin receptor tyrosine kinase substrate p58/53 and the insulin receptor are components of CNS synapses. *J Neurosci* 19: 7300–7308 (1999).
- [25] Florant GL, Singer L, Scheurink AJ, Park CR, Richardson RD and Woods SC. Intraventricular insulin reduces food intake and body weight of marmosets during the summer feeding period. *Physiol Behav* 49: 335–338 (1991).
- [26] Craft S and Watson GS. Insulin and neurodegenerative disease: shared and specific mechanisms. *Lancet Neurol* 3: 169–178 (2004).
- [27] Clarke DD and Sokoloff L. Circulation and energy metabolism of the brain. In: 'Basic neurochemistry'. (Eds: Siegel G J, Agranoff BW, Albers RW, Molinoff SK, Fisher PB, Uhler MD), Lippincott-Ratven, Philadelphia, p. 637–669 (1999).
- [28] Duckrow RB, Beard DC and Brennan RW. Regional cerebral blood flow decreases during chronic and acute hyperglycemia. *Stroke* 18: 52–58 (1987).
- [29] Pelligrino DA, Segil LJ and Albrecht RF. Brain glucose utilization and transport and cortical function in chronic vs. acute hypoglycemia. *Am J Physiol*. 259: E729–E735 (1990).
- [30] Lucignani G, Namba H, Nehlig A, Porrino LJ, Kennedy C and Sokoloff L. Effects of insulin on local cerebral glucose utilization in the rat. *J Cereb Blood Flow Metab* 7: 309–314 (1987).
- [31] Marfaing P, Penicaud L, Broer Y, Mraovitch S, Calando Y and Picon L. Effects of hyperinsulinemia on local cerebral insulin binding and glucose utilization in normoglycemic awake rats. *Neurosci Lett* 115: 279–285 (1990).
- [32] Doyle P, Cusin I, Rohner-Jeanrenaud F and Jeanrenaud B. Four-day hyperinsulinemia in euglycemic conditions alters local cerebral glucose utilization in specific brain nuclei of freely moving rats. *Brain Res* 684(1): 47–55 (1995).
- [33] Fanelli C, Dence C and Markham J. Blood-to-brain glucose transport and cerebral glucose metabolism are not reduced in poorly controlled type 1 diabetes. *Diabetes* 47: 1444–1450 (1998).
- [34] Hasselbalch S, Knudsen G and Videback C. No effect of insulin on glucose blood-brain barrier transport and cerebral metabolism in humans. *Diabetes* 48: 1915–1921 (1999).
- [35] Bingham EM, Hopkins D and Smith D. The role of insulin in human brain glucose metabolism: an 18-fluoro-deoxyglucose positron emission tomography study. *Diabetes* 51: 3384–3390 (2002).
- [36] Brant AM, Jess TJ, Milligan G, Brown CM and Gould GW. Immunological analysis of glucose transporters expressed in different regions of the rat brain and central nervous system. *Biochem Biophys Res Commun* 192: 1297–1302 (1993).
- [37] El Messari S, Leloup C, Quignon M, Brisorgueil MJ, Penicaud L and Arluison M. Immunocytochemical localization of the insulin-responsive glucose transporter 4 (Glut4) in the rat central nervous system. *J Comp Neurol* 399: 492–512 (1998).
- [38] Ibberson M, Uldry M and Thorens B. GLUTX1, a novel mammalian glucose transporter expressed in the central nervous system and insulin-sensitive tissues. *J Biol Chem* 275: 4607–4612 (2000).
- [39] Reagan L, Gorovits N and Hoskin E. Localization and regulation of GLUTx1 glucose transporter in the hippocampus of streptozotocin diabetic rats. *Proc Natl Acad Sci USA* 98:2820–2825 (2001).
- [40] Obici S, Zhang BB, Karkanas G and Rossetti L. Hypothalamic insulin signaling is required for inhibition of glucose production. *Nat Med* 8:1376–82 (2002).
- [41] Niswender KD and Schwartz MW. Insulin and leptin revisited: adiposity signals with overlapping physiological and intracellular signaling capabilities. *Front Neuroendocrinol* 24: 1–10 (2003).
- [42] Clarke DW, Boyd FT Jr, Kappy MS and Raizada MK. Insulin binds to specific receptors and stimulates 2-deoxy-D-glucose uptake in cultured glial cells from rat brain. *J Bio Chem* 259: 11672–11675 (1984).
- [43] Schubert M, Gautam D, Surjo D, Ueki K, Baudler S, Schubert D, *et al.* Role for neuronal insulin resistance in neurodegenerative disease. *Proc Natl Acad Sci USA* 101(9): 3100–3105 (2004).
- [44] Mill JF, Chao MV and Ishii DN. Insulin, insulin-like growth factor II, and nerve growth factor effects on tubulin mRNA levels and neurite formation. *Proc Natl Acad Sci USA* 82: 7126–7130 (1985).
- [45] Wang C, Li Y, Wible B, Angelides KJ and Ishii DN. Effects of insulin and insulin-like growth factors on neurofilament mRNA and tubulin mRNA content in human neuroblastoma SH-SY5Y cells. *Brain Res Mol Brain Res* 13: 289–300 (1992).
- [46] Tanaka M, Sawada M, Yoshida S, Hanaoka F and Marunouchi T. Insulin prevents apoptosis of external granular layer neurons in rat cerebellar slice cultures. *Neurosci Lett* 199: 37–40 (1995).
- [47] Craft S, Asthana S, Newcomer JW, Wilkinson CW, Matos IT, Baker LD, *et al.* Enhancement of memory in Alzheimer's disease with insulin and somatostatin, but not glucose. *Arch Gen Psychiatry* 56: 1135–1140 (1999).

- [48] Kern W, Born J, Schreiber H and Fehm HL. Central nervous system effects of intranasally administered insulin during euglycemia in men. *Diabetes* 48: 557–563 (1999).
- [49] Park CR, Seeley RJ, Craft S and Woods SC. Intracerebroventricular insulin enhances memory in a passive-avoidance task. *Physiol Behav* 68: 509–514 (2000).
- [50] Kern W, Peters A, Fruehwald-Schultes B, Deininger E, Born J and Fehm HL. Improving influence of insulin on cognitive functions in humans. *Neuroendocrinology* 74: 270–280 (2001).
- [51] Craft S, Asthana S, Cook DG, Baker LD, Cherrier M, Purganan K, *et al.* Insulin dose–response effects on memory and plasma amyloid precursor protein in Alzheimer's disease: interactions with apolipoprotein E genotype. *Psychoneuroendocrinology* 28: 809–822 (2003).
- [52] Dou JT, Chen M, Dufour F, Alkon DL and Zhao WQ. Insulin receptor signaling in long-term memory consolidation following spatial learning. *Learn Mem* 12: 646–655 (2005).
- [53] Figlewicz DP, Szot P, Israel PA, Payne C and Dorsa DM. Insulin reduces norepinephrine transporter mRNA *in vivo* in rat locus coeruleus. *Brain Res* 602: 161–164 (1993).
- [54] Kopf S and Baratti C. Effects of posttraining administration of insulin on retention of a habituation response in mice: participation of a central cholinergic mechanism. *Neurobiol Learn Mem* 71: 50–61 (1999).
- [55] Zhao W, Chen H, Xu H, Moore E, Meiri N, Quon MJ, *et al.* Brain insulin receptors and spatial memory. correlated changes in gene expression, tyrosine phosphorylation, and signaling molecules in the hippocampus of water maze trained rats. *J Biol Chem* 274: 34893–34902 (1999).
- [56] Wang YT and Salter MW. Regulation of NMDA receptors by tyrosine kinases and phosphatases. *Nature* 369: 233–5 (1994).
- [57] Wan Q, Xiong ZG, Man HY, Ackerley CA, Braunton J, Lu WY, *et al.* Recruitment of functional GABA(A) receptors to postsynaptic domains by insulin. *Nature* 388: 686–690 (1997).
- [58] Man HY, Lin JW, Ju WH, Ahmadian G, Liu L, Becker LE, *et al.* Regulation of AMPA receptor-mediated synaptic transmission by clathrin-dependent receptor internalization. *Neuron* 25: 649–662 (2000).
- [59] Kneussel M. Dynamic regulation of GABA(A) receptors at synaptic sites. *Brain Res Brain Res Rev* 39: 74–83 (2002).
- [60] Malenka RC. Synaptic plasticity and AMPA receptor trafficking. *Ann NY Acad Sci* 1003: 1–11 (2003).
- [61] Huang CC, Lee CC and Hsu KS. An investigation into signal transduction mechanisms involved in insulin-induced long-term depression in the CA1 region of the hippocampus. *J Neurochem* 89: 217–231 (2004).
- [62] van der Heide LP, Kamal A, Artola A, Gispen WH and Ramakers GM. Insulin modulates hippocampal activity-dependent synaptic plasticity in a N-methyl-d-aspartate receptor and phosphatidylinositol-3-kinase-dependent manner. *J Neurochem* 94: 1158–1166 (2005).
- [63] Ott A, Stolk RP, van Harskamp F, Pols HA, Hofman A and Breteler MM. Diabetes mellitus and the risk of dementia—The Rotterdam Study. *Neurology* 53: 1937–1942 (1999).
- [64] Greenwood C and Winocur G. Glucose treatment reduces memory deficits in young rats fed high-fat diets. *Neurobiol Learn Mem* 75: 179–189 (2001).
- [65] Luchsinger JA, Tang MX, Shea S and Mayeux R. Hyperinsulinemia and risk of Alzheimer disease. *Neurology* 63: 1187–1192 (2004).
- [66] Yaffe K, Blackwell T, Kanaya AM, Davidowitz N, Barrett-Connor E and Krueger K. Diabetes, impaired fasting glucose, and development of cognitive impairment in older women. *Neurology* 63: 658–663 (2004).
- [67] Pirart J. Diabetes mellitus and its degenerative complications—prospective study of 4400 patients observed between 1947 and 1973. *Diab Metab* 3: 245–256 (1977).
- [68] Gispen WH and Biessels GJ. Cognition and synaptic plasticity in diabetes mellitus. *Trends Neurosci* 23: 542–549 (2000).
- [69] Tomlinson DR and Gardiner NJ. Glucose neurotoxicity. *Nat Rev Neurosci* 9(1): 36–45 (2008).
- [70] Ott A, Stolk RP, Hofman A, van Harskamp F, Grobbee DE and Breteler MM. Association of diabetes mellitus and dementia: The Rotterdam study. *Diabetologia* 39: 1392–1397 (1996).
- [71] Leibson CL, Rocca WA, Hanson VA, Cha R, Kokmen E, Obrien PC, *et al.* Risk of dementia among persons with diabetes mellitus: a population-based cohort study. *Am J Epidemiol* 145: 301–308 (1997).
- [72] Fehm HL, Perras B, Smolnik R, Kern W and Born J. Manipulating neuropeptidergic pathways in humans: a novel approach to neuropharmacology?. *Eur J Pharmacol* 405: 43–54 (2000).
- [73] Reger MA, Watson GS, Green PS, Wilkinson CW, Baker LD, Cholerton B, *et al.* Intranasal insulin improves cognition and modulates β -amyloid in early AD. *Neurology* 70(6): 440–448 (2008).
- [74] Schafer M and Erdo SL. Development of glutamate neurotoxicity in cortical cultures: induction of vulnerability by insulin. *Brain Res Dev Brain Res* 62(2): 293–296 (1991).
- [75] Gonzalez C, Farias G and Maccioni RB. Modification of tau to an Alzheimer's type protein interferes with its interaction with microtubules. *Cell Mol Biol (Noisy-le-grand)* 44(7): 1117–1127 (1998).
- [76] Freude S, Plum L, Schnitker J, Leeser U, Udelhoven M, Krone W, *et al.* Peripheral hyperinsulinemia promotes tau phosphorylation *in vivo*. *Diabetes* 54: 3343–3348 (2005).
- [77] Qiu WQ, Ye Z, Kholodenko D, Seubert P and Selkoe DJ. Degradation of amyloid beta-protein by a metalloprotease secreted by microglia and other neural and non-neural cells. *J Biol Chem* 272: 6641–6646 (1997).
- [78] Qiu WQ, Walsh DM, Ye Z, Vekrellis K, Zhang J, Podlisny MB, *et al.* Insulin-degrading enzyme regulates extracellular levels of amyloid beta-protein by degradation. *J Biol Chem* 273: 32730–32738 (1998).
- [79] Gasparini L, Gouras GK, Wang R, Gross RS, Beal MF, Greengard P, *et al.* Stimulation of beta-amyloid precursor protein trafficking by insulin reduces intraneuronal beta-amyloid and requires mitogen-activated protein kinase signaling. *J Neurosci* 21: 2561–2570 (2001).
- [80] Perez A, Morelli L, Cresto JC and Castano EM. Degradation of soluble amyloid beta peptides 1–40, 1–42, and the Dutch variant 1–40Q by insulin degrading enzyme from Alzheimer disease and control brains. *Neurochem Res* 25: 247–255 (2000).
- [81] Farris W, Mansourian S, Chang Y, Lindsley L, Eckman EA, Frosch MP, *et al.* Insulin-degrading enzyme regulates the levels of insulin, amyloid beta-protein, and the beta-amyloid precursor protein intracellular domain *in vivo*. *Proc Natl Acad Sci USA* 100: 4162–4167 (2003).
- [82] Mayeux R, Honig LS, Tang MX, Manly J, Stern Y, Schupf N, *et al.* Plasma A[β]40 and A[β]42 and Alzheimer's disease: relation to age, mortality, and risk. *Neurology* 61: 1185–1190 (2003).
- [83] Cook DG, Leverenz JB, McMillan PJ, Kulstad JJ, Ericksen S, Roth RA, *et al.* Reduced hippocampal insulin-degrading enzyme in late-onset Alzheimer's disease is associated with the apolipoprotein Eepsilon4 allele. *Am J Pathol* 162: 313–319 (2003).
- [84] Ho L, Qin W, Pompl PN, Xiang Z, Wang J, Zhao Z, *et al.* Diet-induced insulin resistance promotes amyloidosis in a transgenic mouse model of Alzheimer's disease. *FASEB J* 18: 902–904 (2004).
- [85] Vanhanen M, Koivisto K, Kuusisto J, Mykkanen L, Helkala EL, Hanninen T, *et al.* Cognitive function in an elderly population with persistent impaired glucose tolerance. *Diabetes Care* 21: 398–402 (1998).
- [86] Wallum BJ, Taborsky GJ Jr, Porte D Jr, Figlewicz DP, Jacobson L, Beard JC, *et al.* Cerebrospinal fluid insulin levels increase during intravenous insulin infusions in man. *J Clin Endocrinol Metab* 64: 190–194 (1987).
- [87] Kaiyala KJ, Prigeon RL, Kahn SE, Woods SC and Schwartz MW. Obesity induced by a high-fat diet is associated with reduced brain insulin transport in dogs. *Diabetes* 49: 1525–1533 (2000).
- [88] Strachan MW, Deary IJ, Ewing FM and Frier BM. Is type II diabetes associated with an increased risk of cognitive dysfunction? A critical review of published studies. *Diabetes Care* 20: 438–445 (1997).
- [89] Caballero AE. Endothelial dysfunction, inflammation, and insulin resistance: a focus on subjects at risk for type 2 diabetes. *Curr Diab Rep* 4: 237–246 (2004).
- [90] Rojo LE, Fernández JA, Maccioni AA, *et al.* Neuroinflammation: implications for the pathogenesis and molecular diagnosis of Alzheimer's disease. *Arch Med Res* 39(1): 1–16 (2008).
- [91] Aggarwal S, Gollapudi S and Gupta S. Increased TNF-alpha-induced apoptosis in lymphocytes from aged humans: changes in TNF-alpha receptor expression and activation of caspases. *J Immunol* 162: 2154–2161 (1999).

- [92] Dzienis-Straczkowska S, Straczkowski M, Szelachowska M, Stepien A, Kowalska I and Kinalska I. Soluble tumor necrosis factor- α receptors in young obese subjects with normal and impaired glucose tolerance. *Diabetes Care* 26: 875–880 (2003).
- [93] Zhao M, Cribbs DH, Anderson AJ, Cummings BJ, Su JH, Wasserman AJ, *et al.* The induction of the TNF α death domain signaling pathway in Alzheimer's disease brain. *Neurochem Res* 28: 307–318 (2003).
- [94] Jagust WJ, Seab JP, Huesman RH, Valk PE, Mathis CA, Reed BR, *et al.* Diminished glucose transport in Alzheimer's disease: dynamic PET studies. *J Cerebr Blood Flow Metab* 11: 323–330 (1991).
- [95] Gibson GE, Petersen C and Jenden DJ. Brain acetylcholine synthesis decline with senescence. *Science* 213: 674–676 (1981).
- [96] Bigl V, Arendt T, Fischer S, Werner M and Arendt A. The cholinergic system in aging. *Gerontology* 33: 172–180 (1987).
- [97] Prapong TJ, Buss WH, Hsu P and Heine H. West Greenlee and E. Uemura. Amyloid beta-peptide decreases neuronal glucose uptake despite causing increase in GLUT3 mRNA transcription and GLUT3 translocation to the plasma membrane. *Exp Neurol* 174: 253–258 (2002).
- [98] Zhu X, Perry G and Smith MA. Insulin signaling, diabetes mellitus and risk of Alzheimer disease. *J Alzheimers Dis* 7: 81–84 (2005).
- [99] Fernandez J, Rojo L, Kuljis RO and Maccioni RB. The danger signals hypothesis of Alzheimer's disease. *J Alz Dis* (2008). In press.
- [100] Moreira PI, Santos MS, Seica R and Oliveira CR. Brain mitochondrial dysfunction as a link between Alzheimer's disease and diabetes. *J Neurol Sci* 257: 206–214 (2007).
- [101] Xie L, Helmerhorst E, Taddei K, Plewright B, Van Bronswijk W and Martins R. Alzheimer's beta-amyloid peptides compete for insulin binding to the insulin receptor. *J Neurosci* 22: RC221 (2002).
- [102] Steen EJ, Terry BJ, Rivera EJ, Cannon JL, Neely TR, Tavares R, *et al.* Impaired insulin and insulin-like growth factor expression and signalling mechanisms in Alzheimer's disease – Is this type 3 diabetes?. *J Alz Dis* 7: 63–80 (2005).
- [103] Rivera EJ, Goldin A, Fulmer N, Tavares R, Wands JR and de la Monte SM. Insulin and insulin-like growth factor expression and function deteriorate with progression of Alzheimer's disease: link to brain reductions in acetylcholine. *J Alzheimers Dis* 8: 247–68 (2005).
- [104] Steven ES, Jongsoo L and Allison BG. Inflammation and insulin resistance. *J Clin Invest* 116(7): 1793–1801 (2006).
- [105] Röder HM and Ingram VM. Two novel kinases phosphorylate tau and the KSP site of heavy neurofilament subunits in high stoichiometric ratios. *J Neurosci* 11: 3325–3342 (1991).
- [106] Bush ML, Miyashiro JS and Ingram VM. Activation of a neurofilament kinase, a tau kinase, and a tau phosphatase by decreased ATP levels in nerve growth factor-differentiated PC12 cells. *Proc Natl Acad Sci USA* 92: 1962–1965 (1995).
- [107] Hong M and Lee VM. Insulin and insulin-like growth factor-1 regulate tau phosphorylation in cultured human neurons. *J Biol Chem* 272: 19547–19553 (1997).
- [108] Rojo L, Sjöberg MK, Hernández P, Zambrano C and Maccioni RB. Roles of cholesterol and lipids in the etiopathogenesis of Alzheimer's disease. *J Biomed Biotechnol* 2006(3): 73976 (2006).
- [109] Gasparini L, Netzer WJ, Greengard P and Xu H. Does insulin dysfunction play a role in Alzheimer's disease?. *Trends Pharmacol Sci* 23: 288–293 (2002).
- [110] Cross DA, Alessi DR, Cohen P, Andjelkovich M, Hemmings BA. Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature* 378: 785–789 (1995).
- [111] Fisher TL and White MF. Signaling pathways: the benefits of good communication. *Current Biology* 14: R1005–R1007 (2004).
- [112] Dudek H, Datta SR, Franke TF, Birnbaum MJ, Yao R, Cooper GM, *et al.* Regulation of neuronal survival by the serine-threonine protein kinase Akt. *Science* 275: 661–665 (1997).
- [113] Zhou H, Summers SA, Birnbaum MJ and Pittman RN. Inhibition of Akt kinase by cell-permeable ceramide and its implications for ceramide-induced apoptosis. *J Biol Chem* 273: 16568–16575 (1998).
- [114] Zhou H, Li XM, Meinkoth J and Pittman RN. Akt regulates cell survival and apoptosis at a postmitochondrial level. *J Cell Biol* 151: 483–494 (2000).
- [115] Martin D, Salinas M, Lopez-Valdaliso R, Serrano E, Recuero M and Cuadrado A. Effect of the Alzheimer amyloid fragment Abeta(25–35) on Akt/PKB kinase and survival of PC12 cells. *J Neurochem* 78: 1000–1008 (2001).
- [116] Maccioni RB, Otth C, Concha II and Muñoz JP. The protein kinase Cdk5. Structural aspects, roles in neurogenesis and involvement in Alzheimer's pathology. *Eur J Biochem* 268(6): 1518–1527 (2001).
- [117] Alvarez A, Toro R, Caceres A, Maccioni RB. Inhibition of tau phosphorylating protein kinase cdk5 prevents beta-amyloid-induced neuronal death. *FEBS Lett* 459(3): 421–426 (1999).
- [118] Alvarez A, Muñoz JP and Maccioni RB. A Cdk5–p35 Stable Complex Is Involved in the β -Amyloid-Induced Deregulation of Cdk5 Activity in Hippocampal Neurons. *Exp Cell Res* 264(2): 266–274 (2001).
- [119] Shiurba RA, Ishiguro K, Takahashi M, Sato K, Spooner ET, Mercken M, *et al.* Immunocytochemistry of tau phosphoserine 413 and tau protein kinase I in Alzheimer pathology. *Brain Res* 737(1–2): 119–132 (1996).
- [120] Leroy K, Boutajangout A, Authelat M, Woodgett JR, Anderton BH and Brion JP. The active form of glycogen synthase kinase-3 β is associated with granulovacuolar degeneration in neurons in Alzheimer's disease. *Acta Neuropathol (Berl)* 103: 91–99 (2002).
- [121] Ryu BR, Ko HW, Jou I, Noh JS and Gwag BJ. Phosphatidylinositol 3-kinase-mediated regulation of neuronal apoptosis and necrosis by insulin and IGF-I. *J Neurobiol* 39: 536–546 (1999).
- [122] Philpott KL, McCarthy MJ, Klippel A and Rubin LL. Activated phosphatidylinositol 3-kinase and Akt kinase promote survival of superior cervical neurons. *J Cell Biol* 139: 809–815 (1997).
- [123] Matsuzaki H, Tamatani M, Mitsuda N, Namikawa K, Kiyama H, Miyake S, *et al.* Activation of Akt kinase inhibits apoptosis and changes in Bcl-2 and Bax expression induced by nitric oxide in primary hippocampal neurons. *J Neurochem* 73: 2037–2046 (1999).
- [124] Yamaguchi A, Tamatani M, Matsuzaki H, Namikawa K, Kiyama H, Vitek MP, *et al.* Akt activation protects hippocampal neurons from apoptosis by inhibiting transcriptional activity of p53. *J Biol Chem* 276: 5256–5264 (2001).
- [125] Sanna PP, Cammalleri M, Berton F, Simpson C, Lutjens R, Bloom FE, *et al.* Phosphatidylinositol 3-kinase is required for the expression but not for the induction or the maintenance of long-term potentiation in the hippocampal CA1 region. *J Neurosci* 22: 3359–3365 (2002).
- [126] Watson GS and Craft S. The role of insulin resistance in the pathogenesis of Alzheimer's disease: implications for treatment. *CNS Drugs* 17(1): 27–45 (2003).
- [127] Craft S. Insulin resistance syndrome and Alzheimer's disease: Age- and obesity-related effects on memory, amyloid, and inflammation. *Neurobiol Aging* 26S: S65–S69 (2005).
- [128] Craft S. Insulin resistance syndrome and Alzheimer disease: pathophysiologic mechanisms and therapeutic implications. *Alzheimer Dis Assoc Disord* 20: 298–301 (2006).
- [129] Watson GS, Cholerton BA, Reger MA, Baker LD, Plymate SR, Asthana S, *et al.* Preserved cognition in patients with early Alzheimer disease and amnesic mild cognitive impairment during treatment with rosiglitazone: a preliminary study. *Am J Geriatr Psychiatry* 13: 950–958 (2005).
- [130] Hoyer S. Glucose metabolism and insulin receptor signal transduction in Alzheimer disease. *Eur J Pharmacol* 490(1–3): 115–125 (2004).
- [131] Reger MA, Watson GS, Frey II WH, Baker L, Cholerton B, Keeling M, *et al.* Effects of intranasal insulin on cognition in memory-impaired older adults: Modulation by APOE genotype. *Neurobiol Aging* 27: 451–458 (2006).
- [132] Jiang Q, Heneka M and Landreth GE. The Role of Peroxisome Proliferator-Activated Receptor-gamma (PPAR γ) in Alzheimer's Disease: Therapeutic Implications. *CNS Drugs* 22(1): 1–14 (2008).
- [133] Sastre M, Dewachter I, Landreth GE, Willson TM, Klockgether T, van Leuven F and Heneka MT. Nonsteroidal anti-inflammatory drugs and peroxisome proliferator-activated receptor gamma agonists modulate immunostimulated processing of amyloid precursor protein through regulation of beta-secretase. *J Neurosci* 23: 9796–9804 (2003).
- [134] Camacho IE, Serneels I, Spittaels K, Merchiers P, Dominguez D and De Strooper B. Peroxisome-proliferator-activated receptor gamma induces a clearance mechanism for the amyloid-beta peptide. *J Neurosci* 24: 10908–10917 (2004).

- [135] d'Abramo C, Massone S, Zingg JM, Pizzuti A, Marambaud P, Dalla Piccola B, *et al.* Role of peroxisome proliferator-activated receptor gamma in amyloid precursor protein processing and amyloid beta-mediated cell death. *Biochem J* 391: 693–698 (2005).
- [136] d'Abramo C, Ricciarelli R, Pronzato, MA and Davis P. Troglitazone, a peroxisome proliferator-activated receptor-c agonist, decreases tau phosphorylation in CHOtau4R cells. *J Neurochem* 98: 1068–1077 (2006).
- [137] Florez JC, Jablonski KA, Sun MW, Bayley N, Kahn SE, Shamooh H, *et al.* Effects of the type 2 diabetes-associated PPAR γ Pro12Ala polymorphism on progression to diabetes and response to troglitazone. *J Clin Endocrinol Metab* 92: 1502–1509 (2007).
- [138] Türkmen Kemal Y, Güvener Demirag N, Yildirir A, Atar A, Dogruk Unal A and Biyiklioglu Z. Effects of rosiglitazone on plasma brain natriuretic peptide levels and myocardial performance index in patients with type 2 diabetes mellitus. *Acta Diabetol* 44(3): 149–156 (2007).
- [139] Nissen SE and Wolski K. Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular disease. *N Engl J Med* 356: 2457–2471 (2007).
- [140] McGuire D and Inzucchi S. New Drugs for the Treatment of Diabetes Mellitus Part I: Thiazolidinediones and Their Evolving Cardiovascular Implications. *Circulation* 117: 440–449 (2008).
- [141] Risner M, Saunders A, Altman J, Ormandy G, Craft S, Foley I, *et al.* Efficacy of rosiglitazone in a genetically defined population with mild-to-moderate Alzheimer's disease. *Pharmacogenomics J* 6: 246–254 (2006).
- [142] Lavados M, Farias G, Rothhammer F, Guillon M, Mujica MC, Maccioni C, *et al.* ApoE alleles and tau markers in patients with different levels of cognitive impairment. *Arch Med Res* 36: 474–479 (2005).

Received: ?????????????? Revised: ?????????????? Accepted: ??????????????